



Effect of Quercetin on the Activity and mRNA Expression of Antioxidant Enzymes and Physiological Responses in Olive Flounder (*Paralichthys olivaceus*) Exposed to Cadmium*

H. S. Shin, J. H. Yoo¹, T. S. Min², J. Lee³ and C. Y. Choi**

Division of Marine Environment and Bioscience, Korea Maritime University, Busan 606-791, Korea

ABSTRACT : We investigated the antioxidant efficacy of quercetin (0% Diet 1, 0.25% Diet 2, and 0.5% Diet 3) pretreatment for 30 and 60 days in response to cadmium (Cd) toxicity in the olive flounder, and measured the plasma lysozyme activity to understand the immune effects of quercetin. The lysozyme activity with Diets 2 and 3 was higher than with Diet 1. Based on this result, to examine the immune ability and antioxidant role of quercetin, we exposed olive flounder fed quercetin to Cd and then measured the expression and activity of antioxidant enzymes (superoxide dismutase (SOD) and catalase (CAT)) and lipid peroxidation (LPO). With Diets 2 and 3, the expression and activity of antioxidant enzymes and the H₂O₂ concentration were lower than with Diet 1. In addition, the LPO levels were lower than with Diet 1, which protected the cell membrane. Therefore, quercetin removed the reactive oxygen species (ROS) produced by Cd, indicating that quercetin has antioxidant ability. In addition to its antioxidant ability, quercetin has immune effects. (**Key Words :** Antioxidant, LPO, Lysozyme, Quercetin, ROS, Cadmium, Olive Flounder)

INTRODUCTION

Many environmental stress such as cadmium (Cd) increase reactive oxygen species (ROS) and provoke oxidative stress in organisms (Storey, 1996). ROS include superoxide (O₂⁻), hydrogen peroxide (H₂O₂), the hydroxyl radical (HO[·]), and singlet oxygen (¹O₂) (Kinnula et al., 1995). Overproduction of ROS by Cd can increase lipid peroxidation (LPO), the oxidation of nucleic acid and proteins, and DNA damage.

To protect themselves against ROS generating oxidative stress, aerobic organisms have evolved complex antioxidant defense systems. Antioxidant enzymes such as superoxide

dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX), and glutathione S-transferase (GST), and low-molecular-weight antioxidant materials such as vitamin C and vitamin E are found in the livers and kidneys of marine organisms (Basha and Rani, 2003; Hansen et al., 2006).

Previous studies (Pinho et al., 2005; Weng et al., 2007; Prieto et al., 2008) have examined systems involving natural antioxidants. These studies investigated the antioxidant effects of vitamin E supplements in the crab (*Chasmagnathus granulatus*) (Pinho et al., 2005), mouse (*Mus musculus*) (Weng et al., 2007), and tilapia (*Oreochromis niloticus*) (Prieto et al., 2008) against microcystin toxicity by analyzing CAT and GST activity. In isolated carp (*Cyprinus carpio* var. *Jian*) enterocytes, Chen et al. (2009) showed that the antioxidant effect of glutamine scavenging of ROS induced by H₂O₂ treatment occurred via the activity of the antioxidant enzymes SOD and CAT.

In this study investigated the effect of the natural antioxidant quercetin in olive flounder exposed to Cd. Quercetin is a polyphenolic flavonoid compound that is a strong antioxidant and is almost ubiquitous in plants. Broccoli and apples contain 7-110 mg/kg, while onions contain 284-486 mg/kg (Scalbert and Williamson, 2000). Quercetin chelates metal ions, has free radical scavenging activity, and reduces the concentration of metal to protect

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** Corresponding Author: C. Y. Choi. Tel: +82-51-410-4756, Fax: +82-51-404-4750, E-mail: choic@hhu.ac.kr

¹ Jeilfeed company Ltd., Kyoungnam 637-833, Korea.

² Division of Medical and Pharmaceutical Science, National Research Foundation of Korea, Daejeon 305-350, Korea.

³ Department of Marine Life Sciences, Jeju National University, Jeju Special Self-Governing Province, 690-756, Korea.

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cell membranes (Bors and Saran, 1987). It can stop redox reactions by chelating ROS generated by toxic materials like Cd, and it inhibits LPO on the cell membrane. The phenoxy radical generated combines with another ROS and then inhibits the production of ROS (Frankle et al., 1993).

Many recent studies have examined the effects of antioxidants against oxidative stress induced by toxic materials. For example, Jayaraj et al. (2007) investigated the effects of pretreatment with three flavonoids (silybin, quercetin, and morin) on the effects of microcystins. Many studies of a variety of antioxidants have been reported, including vitamin C (Qinghui et al., 2004; Weng et al., 2007), lycopene (Al-Jassabi, 2005), and tea polyphenols (Xu et al., 2007). In fish, although many studies have examined how supplementation with vitamin C (Trenzado et al., 2009), vitamin E (Prieto et al., 2008), selenium (Atencio et al., 2009), and glutamine (Chen et al., 2009) affect the antioxidant system, little is known about the antioxidant effects of supplemental quercetin.

Antioxidant improved immune effect as well as antioxidant ability. Qinghui et al. (2004) reported beneficial effects of vitamin C on immunological parameters, such as lysozyme, in the Japanese seabass (*Lateolabrax japonicus*). Increased immune ability was demonstrated by the increased lysozyme (Ortuno et al., 1999; Ai et al., 2004). Many antioxidants are reported to enhance immune capacity as well as antioxidant capacity.

Therefore, this study determined the effects of quercetin on enhancing growth and immune ability by measuring the plasma lysozyme activity. Then, we investigated the effect of quercetin pretreatment on the toxicity induced by Cd in the olive flounder by measuring the expression and activity of antioxidant enzymes (SOD and CAT), the plasma H₂O₂ concentration, and LPO as an oxidative stress parameter.

MATERIALS AND METHODS

Experimental fish and conditions

Olive flounders (n = 800, length 10±0.5 cm, weight 19.9±1.3 g) were obtained from a commercial fish farm and acclimated to the experimental conditions for 2 weeks in nine 300 L circulation filter tanks in the laboratory. During the experiments, the water temperature and photoperiod were maintained at 20±1°C and a 12L:12D cycle, respectively. The fish were fed a commercial and experimental diets feed twice daily (09:00 and 17:00). In all, 800 flounders were chosen randomly and distributed in the nine 300 L flow-through tanks.

Extraction of quercetin

The extraction of quercetin was carried out according to the methods of Velioglu and Mazza (1991) and Kang et al.

(1998), with modification. The dried onion peels were blended with methanol in a Waring blender for 5 min and filtered through Whatman No. 1 filter paper, and then the filtrate was concentrated using a rotary evaporator at 40°C. The residue was washed with ether in a separatory funnel to remove lipids and other fat soluble materials. The onion extract was fractionated in the order of ethylacetate to purified glucosides type of quercetin, and then we examined quercetin content level by HPLC.

Experimental diets

Fish meal, dehulled soybean, and corn gluten meal were used as protein sources, and wheat flour and squid liver oil were used as carbohydrate and lipid sources, respectively. The ingredients of the experimental diets were mixed well with water in a ratio of 3:1 and then pelletized. The experiment diets were dried at room temperature and stored at -20°C until required. In the experimental diets, the crude protein content ranged from 55.1-56.0%, and the crude lipid content from 9.1-9.7%. The estimated energy content was 4.1 kcal/g.

Experimental diets using purified quercetin were made by jeilfeed company (Kyoungnam, Korea), and were coated for protecting soluble nutrients include quercetin in the water. Experimental diets were in the water for a while, quercetin content level was not significantly different from the content of original experimental diets. The experimental diets contained 0% (Diet 1), 0.25% (Diet 2), or 0.5% (Diet 3) quercetin, at the expense of 0, 0.25, or 0.5% wheat flour, respectively (Table 1).

Plasma lysozyme activity

To determine the lysozyme activity of olive flounder, 50 µl of plasma were added to 950 µl of a suspension of *Micrococcus lysodeikticus* (0.2 mg/ml) in 0.05 M sodium phosphate buffer (pH 6.2). The reactions were carried out at 25°C, and the absorbance at 530 nm was measured between 0.5 and 4.5 min. One lysozyme activity unit was defined as the amount of enzyme producing a 0.001/min decrease in absorbance.

Cd exposure

The experimental fish were exposed to CdCl₂·2.5H₂O (Kanto Chemical, Tokyo, Japan) dissolved in water to a Cd²⁺ concentration of 10 ppb in 50 L tanks for 0, 6, 12, 24, and 48 h. Four fish from each group (Diets 1, 2, and 3) were selected randomly for blood and tissue sampling after fed Diet 1, 2 and 3 for 30 and 60 days respectively. The fish were anesthetized with 200 mg/L tricaine methanesulfonate (MS-222, Sigma, USA) before blood collection. Blood was collected from the caudal vein using a 3-ml syringe coated with heparin. Plasma samples were separated by

Table 1. Ingredients and nutrient composition of the experimental diets

Ingredient (%)	Diet		
	Diet 1 (0%)	Diet 2 (0.25%)	Diet 3 (0.5%)
Fish meal ¹ (ML)	45.00	45.00	45.00
Corn gluten ML	15.00	15.00	15.00
Wheat flour	19.90	19.65	19.40
Soybean ML	10.00	10.00	10.00
Fish oil - salmon	2.50	2.50	2.50
Squid ML	2.00	2.00	2.00
Krill ML	2.00	2.00	2.00
Lecithin	1.00	1.00	1.00
Mono-calcium	1.00	1.00	1.00
Choline	1.00	1.00	1.00
VITAMIX ²	0.20	0.20	0.20
MINEMIX ³	0.20	0.20	0.20
Vitamin C	0.20	0.20	0.20
QUERCETIN	0.00	0.25	0.50
Total	100	100	100

¹ Imported from Chile.

² Vitamin premix contained the following ingredients (g/kg mix): L-ascorbic acid, 121.2; DL- α -tocopheryl acetate, 18.8; thiamin hydrochloride, 2.7; riboflavin, 9.1; pyridoxine hydrochloride, 1.8; niacin, 36.4; Ca-D-pantothenate, 12.7; myo-inositol, 181.8; D-biotin, 0.27; folic acid, 0.68; p-aminobenzoic acid, 18.2; menadione, 1.8; retinyl acetate, 0.73; cholecalciferol, 0.003; cyanocobalamin, 0.003.

³ Mineral premix contained the following ingredients (g/kg mix): MgSO₄·7H₂O, 80.0; NaH₂PO₄·2H₂O, 370.0; KCl, 130.0; Ferric citrate, 40.0; ZnSO₄·7H₂O, 20.0; Ca-lactate, 356.5; CuCl, 0.2; AlCl₃·6H₂O, 0.15; KI, 0.15; Na₂Se₂O₃, 0.01; MnSO₄·H₂O, 2.0; CoCl₂·6H₂O, 1.0.

centrifugation (4°C, 10,000 g, 5 min), and stored at -80°C until analysis. To collect liver tissue samples, fish were euthanized by spinal transection. Immediately after collection, the samples were frozen in liquid nitrogen and stored at -80°C until total RNA was extracted for analysis.

Quantitative PCR (QPCR)

QPCR was conducted to determine the relative expression of SOD and CAT mRNA in total RNA extracted from the liver. Primers for QPCR were designed with reference to the known sequences of olive flounder as follows: SOD forward primer (5'-CGT TGG AGA CCT GGG GAA TGT G-3'), SOD reverse primer (5'-ATC GTC AGC CTT CTC GTG GAT C-3'), CAT forward primer (5'-GGC TGA GAA GTT CCA GTT CAA TCC-3'), CAT reverse primer (5'-CTC CAC CTC TGC AAA GTA GTT GAC-3'), β -actin forward primer (5'-GCA AGA GAG GTA TCC TGA CC-3'), and β -actin reverse primer (5'-CTC AGC TCG TTG TAG AAG G-3'). PCR amplification was conducted using a BIO-RAD iCycler iQ Multicolor Real-Time PCR Detection System (Bio-Rad, USA) and iQTM SYBR Green Supermix (Bio-Rad, USA) according to the

manufacturer's instructions. QPCR was performed as follows: denaturation at 95°C for 5 min; 35 cycles of denaturation at 95°C for 20 s, annealing at 55°C for 20 s. As an internal control, experiments were duplicated with β -actin, and all data were expressed as change with respect to corresponding β -actin calculated threshold cycle (CT) levels.

SOD and CAT activity analysis

Tissues were homogenized in ice-cold 0.1 M phosphate-buffered saline (PBS, pH 7.4) containing 1 mM EDTA. The homogenates were centrifuged at 10,000 g for 15 min at 4°C, the supernatant was removed, and the remaining sample was analyzed. SOD and CAT activities were determined using commercial kits supplied by Cayman Chemical (USA).

The SOD activity was assessed using a tetrazolium salt to detect superoxide radicals generated by xanthine oxidase and hypoxanthine. One unit of SOD is defined as the amount of enzyme needed to exhibit 50% dismutation of the superoxide radical. The absorbance is read at 450 nm. Each assay was performed in duplicate, and enzyme units were recorded as U/ml.

The method for CAT activity is based on the reaction of the enzyme with methanol in the presence of an optimal concentration of H₂O₂. The formaldehyde produced is measured spectrophotometrically with 4-amino-3-hydrazino-5-mercapto-1,2,4-triazole (Purpald) as the chromogen (Wheeler et al., 1990). Purpald specifically forms a bicyclic heterocycle with aldehydes and changes from colorless to purple on oxidation. The absorbance is read at 540 nm. Each assay was performed in duplicate, and the CAT activity was expressed as nmol/min/ml.

H₂O₂ assay

H₂O₂ concentrations were measured using the modified methods of Nouroozzadeh et al. (1994) and a Peroxidetect kit (Sigma, USA). 20 μ l of olive flounder serum was added per well to flat bottom 96-well microtitre plates. Plates were left at room temperature for 20 min to allow serum to settle and adhere. A working color reagent was prepared by mixing 100 ml distilled water containing 100 mM sorbitol and 125 μ M xylenol orange (Sigma, USA) with 1 ml of 25 mM ferrous ammonium sulphate prepared in 2.5 M sulphuric acid (Sigma-Aldrich). 200 μ l of this reagent was then added to each well and allowed to incubate at room temperature for 1 h. Absorbance was read at 560 nm and concentration of H₂O₂ were interpolated from a standard curve. Concentrations are expressed as nmole peroxide/ml.

LPO assay

LPO is quantified by measuring malondialdehyde

(MDA) and 4-hydroxynonenal (4-HNE), the degradation products of polyunsaturated fatty acids (PUFAs) hydroperoxides (Esterbauer et al., 1991). The Cayman Chemical (Ann Arbor, MI, USA) lipid hydroperoxide assay kit was used to measure hydroperoxides directly, utilizing the redox reaction with ferrous ion. Hydroperoxides were extracted into chloroform and reacted with ferrous ions to produce ferric ions. The resulting ferric ions were detected using thiocyanate ion as the chromogen. The hydroperoxide concentration was determined based on the absorption at 500 nm.

Statistical analysis

All data were analyzed using the SPSS statistical package (version 10.0; SPSS Inc., USA). One way ANOVA followed by *post hoc* Duncan's multiple range test was used to compare the differences in the data ($p < 0.05$).

RESULTS

Quercetin contents

The amount of quercetin (glucosides type) from onion by HPLC was about 1 g/kg onion (0.1%), and then formulated to contain three concentrations (0, 0.25, and 0.5%) of extracted quercetin by replacing wheat flour in experimental diets.

Growth performance

The weight gain of flounder fed Diet 3, which contained quercetin, for 30 and 60 days were significantly higher than those in flounder fed Diet 1, which did not contain quercetin. This result indicated that high concentration (Diet 3) of quercetin is very effective than low concentration (Diet 2) in growth (Figure 1).

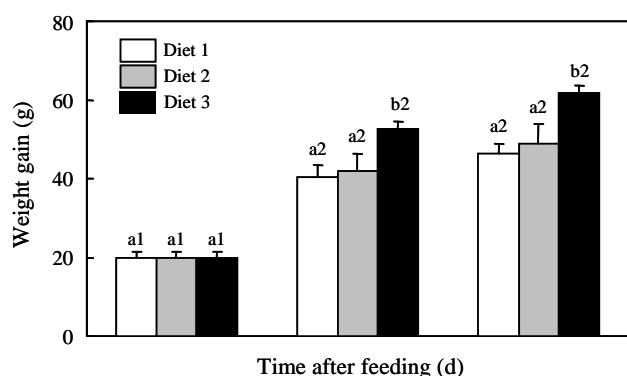


Figure 1. Weight gain of olive flounder fed experimental diets containing quercetin. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after feeding. The numbers indicate significant differences from the initial weight (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

Plasma lysozyme activity

In Diets 2 and 3, the lysozyme activity was significantly higher than in Diet 1 after feeding flounder quercetin for 30 and 60 days. In addition, the lysozyme activities in Diets 1, 2, and 3 fed for 60 days were significantly higher than were those fed for 30 days (Figure 2).

QPCR for SOD and CAT mRNA expression

Using a quantitative polymerase chain reaction (QPCR), we examined the changes in SOD and CAT mRNA expression when olive flounder fed a diet containing quercetin were exposed to Cd. After feeding for 30 days, for Diet 1, the SOD mRNA expression was increased significantly at 24 h and then decreased, whereas the expression levels with Diets 2 and 3 were lower than with Diet 1 (Figure 3A); the CAT mRNA was increased at 6 h and then decreased, whereas the expression levels with Diets 2 and 3 were lower than with Diet 1 (Figure 3C). After feeding for 60 days, with Diet 1, the SOD mRNA expression was increased significantly at 12 h and then decreased, whereas the expression levels with Diets 2 and 3 were lower than with Diet 1 (Figure 3B); the CAT mRNA was increased at 12 h and then decreased, whereas the expression levels with Diets 2 and 3 were lower than with Diet 1 (Figure 3D).

SOD and CAT activity

The effects of quercetin on Cd-induced antioxidant enzyme (SOD and CAT) activities are shown in Figure 3. After feeding for 30 days, with Diet 1, the SOD activity was increased significantly at 12 h and then decreased, whereas the activity levels with Diets 2 and 3 were increased significantly at 6 and 12 h and then decreased, but they were lower than with Diet 1 (Figure 4A); the CAT activity

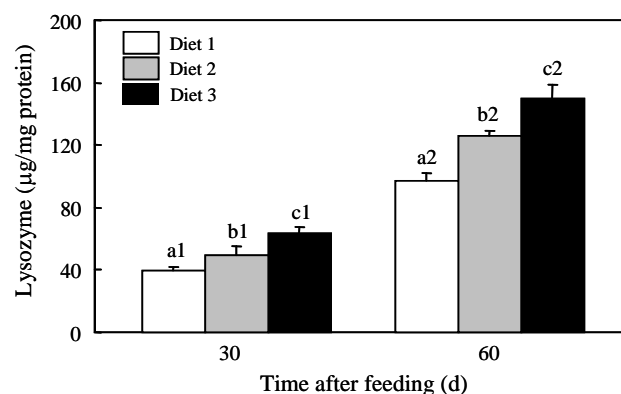


Figure 2. Lysozyme activity in plasma of olive flounder fed diets containing 0, 0.25, and 0.5% quercetin for 30 and 60 days. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after feeding. The numbers indicate significant differences from the 30 days (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

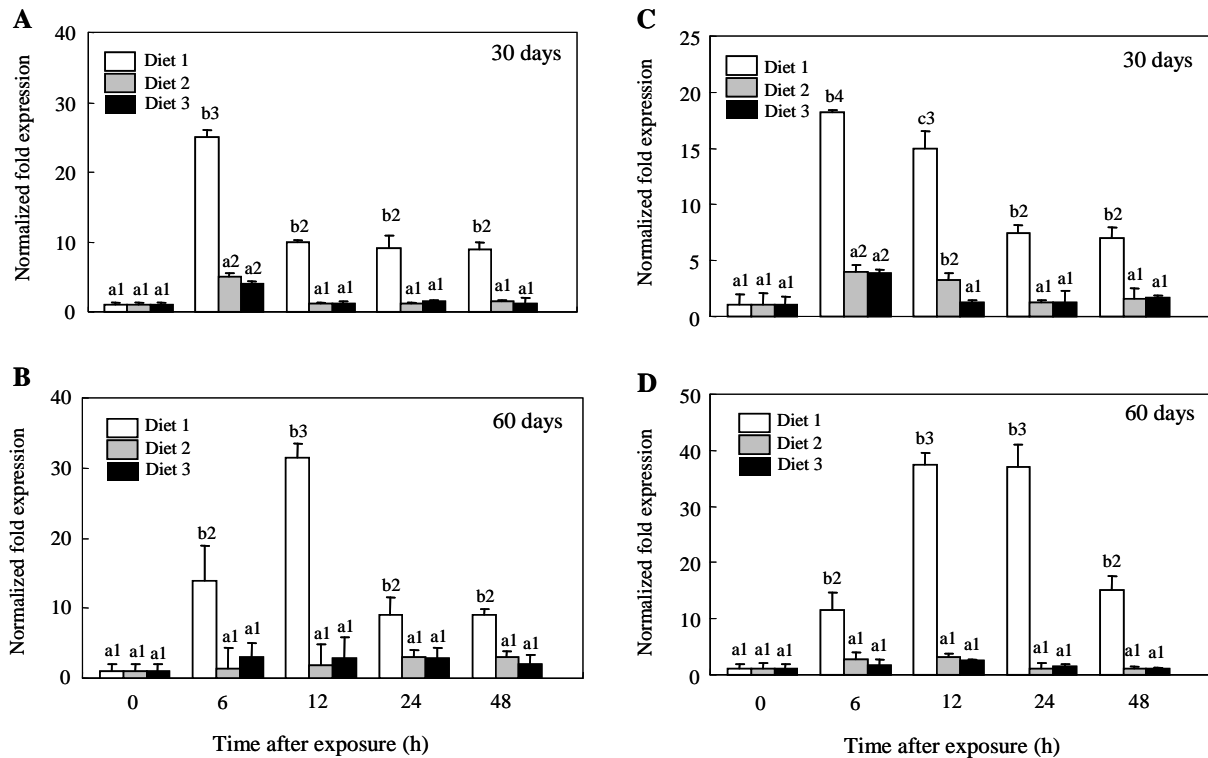


Figure 3. Expression of SOD (A) and CAT (C) in olive flounder fed a diet containing quercetin for 30 days and SOD (B) and CAT (D) in olive flounder fed a diet containing quercetin for 60 days. The mRNA levels in the livers of olive flounder exposed to Cd (0, 6, 12, 24, and 48 h) determined using quantitative real-time PCR. First, 2.5 μ g of total RNA prepared from the liver was reverse-transcribed and amplified using gene-specific primers. The results are expressed as normalized fold expressions with respect to β -actin levels for the same sample. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after exposure. The numbers indicate a significant difference from the control (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

was increased at 12 h and maintained until 48 h, and the activity level with Diets 2 and 3 was increased significantly at 12 h and then decreased, but they were lower than with Diet 1 (Figure 4C). After feeding for 60 days, with Diet 1, the SOD activity was increased significantly at 6 h and maintained until 48 h, and the activity levels with Diets 2 and 3 were increased significantly at 12 h and then decreased, but they were lower than with Diet 1 (Figure 4B); the CAT activity increased gradually until 48 h, and the activity levels with Diet 2 and 3 were increased significantly at 12 h and then decreased, but they were lower than with Diet 1 (Figure 4D).

H₂O₂ assay

After feeding for 30 days, with Diet 1, the plasma H₂O₂ concentration increased significantly at 6 h and then decreased, and H₂O₂ concentrations with Diet 2 and 3 were increased significantly at 12 h and then decreased to initial levels, but they were lower than with Diet 1. After feeding for 60 days, with Diet 1, the plasma H₂O₂ concentration increased significantly until 24 h and then decreased, and the H₂O₂ concentrations with Diet 2 and 3 were lower than with Diet 1 and not significantly different from the initial levels (Figure 5).

LPO assay

After feeding for 30 days, with Diet 1, the LPO level (expressed as MDA and 4-HNE compounds) increased significantly until 48 h, and the LPO levels with Diets 2 and 3 were not significantly different from the initial levels, but they were lower than with Diet 1. After feeding for 60 days, with Diet 1, the LPO level increased significantly at 24 h; with Diet 2, the LPO level increased significantly at 6 h and was maintained until 48 h; and with Diet 3, no significant difference was observed compared with the initial levels. The levels with Diets 2 and 3 were lower than with Diet 1 (Figure 6).

DISCUSSION

To understand the antioxidant effect of quercetin on oxidative stress induced by Cd in olive flounder, we investigated the effects on lysozyme, and the expression and activity of SOD and CAT.

The weight gain of flounder fed Diet 3 for 30 and 60 days were significantly higher than those in flounder fed Diet 1 and 2 (Figure 1). This result indicated that high concentration (Diet 3) of quercetin is very effective than

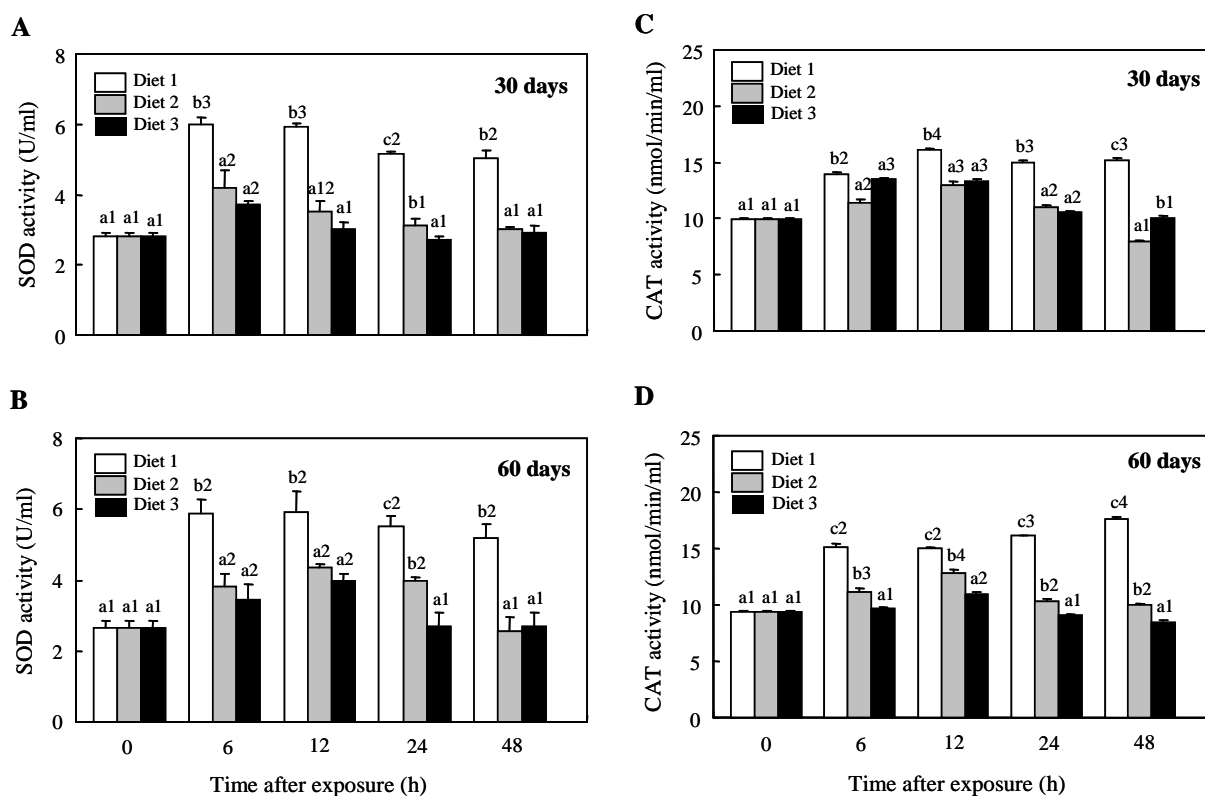


Figure 4. Activity of SOD (A) and CAT (C) in olive flounder fed a diet containing quercetin for 30 days and SOD (B) and CAT (D) in olive flounder fed a diet containing quercetin for 60 days. These activities in livers of olive flounder exposed to Cd (0, 6, 12, 24, and 48 h) determined using a microplate reader. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after exposure. The numbers indicate a significant difference from the control (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

low concentration (Diet 2) in growth. Antioxidant, vitamin C could enhance the growth in fresh shrimp (*Penaeus chinensis*) (Wang and Li, 1996), African catfish (*Clarias gariepinus*) (Merchie et al., 1997a) and carp (*Cyprinus carpio*) (Gouillou-Coustans et al., 1998).

The plasma lysozyme activity with Diets 2 and 3 were significantly higher than with Diet 1 after feeding for 30 or 60 days (Figure 2). These results concur with those of Eo and Lee (2008), who reported that vitamin C improved the immune responses and disease resistance by increasing plasma lysozyme activity in Tiger puffer (*Takifugu rubripes*) fed vitamin C for 8 weeks, and with Zheng et al. (2009), who observed higher lysozyme activity in channel catfish (*Ictalurus punctatus*) fed oregano as an antioxidant.

The expression of SOD and CAT mRNA with Diets 2 and 3 was significantly lower than with Diet 1 (Figure 3). Similarly, the SOD and CAT activities were increased significantly in all groups exposed to Cd, but the activities with Diets 2 and 3 were significantly lower than with Diet 1 (Figure 4). These results are similar to those of Jayaraj et al. (2007), who reported that the CAT activity in mice exposed to microcystin after feeding with quercetin was significantly lower than in controls, suggesting that quercetin, with its

strong antioxidant activity, protects hepatic cells from toxic materials that induce oxidative stress. In addition, the activities of SOD and CAT in a freshwater fish, the matrinxã (*Brycon cephalus*), exposed to methyl parathion after feeding selenium were significantly lower than in controls, indicating that selenium is an antioxidant that scavenges the ROS induced by methyl parathion (Monteiro et al., 2009). Combined with the results of previous studies, the fact that the expression and activity of antioxidant enzymes in the fish fed Diets 2 and 3 were lower than in those fed Diet 1 indicates that quercetin has antioxidant activity by scavenging the ROS induced by Cd directly. In addition, the H_2O_2 concentrations with Diets 2 and 3 were significantly lower than with Diet 1 (Figure 5), also indicating that quercetin scavenges ROS overproduced in the olive flounder.

The oxidative stress caused by ROS generates LPO and damages cells (Valavanidis et al., 2006). In this study, the LPO levels with Diets 2 and 3 were significantly lower than with Diet 1 (Figure 6). These results agree with Hiratsuka et al. (2008), who reported that LPO levels were reduced in mice fed docosahexaenoic acid (DHA), indicating that antioxidant can protect biomembranes and lipid

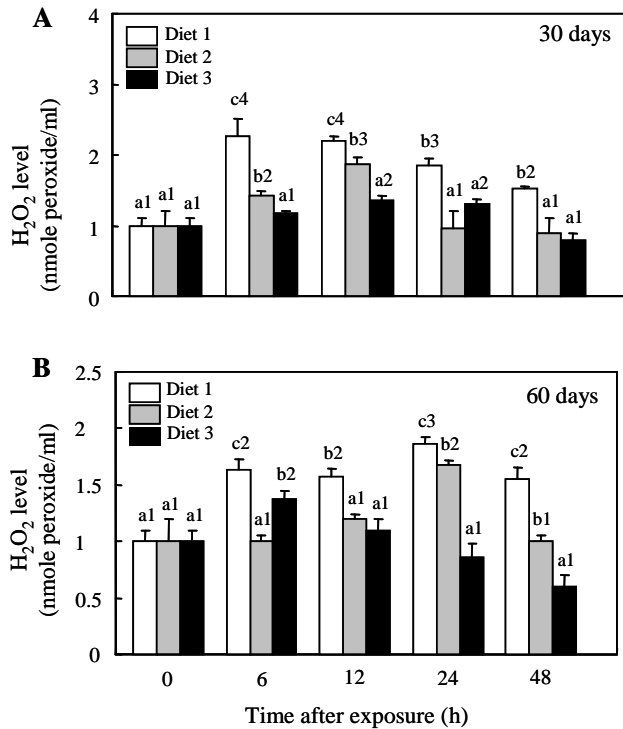


Figure 5. H₂O₂ concentrations in plasma of olive flounder fed for 30 or 60 days diets containing quercetin 0.25 or 0.5%, after 6, 12, 24, and 48 h exposure to Cd. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after exposure. The numbers indicate a significant difference from the control (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

components containing unsaturated fatty acids against the attack from ROS.

In conclusion, quercetin increases lysozyme activity enhancing immune ability in olive flounder. In addition, the mRNA expression and activity of the antioxidant enzymes SOD and CAT and the H₂O₂ concentrations in fish fed Diets 2 and 3 were significantly lower with Diet 1, indicating that quercetin scavenges the ROS induced by Cd to enhance antioxidant effects. Hence, we confirmed that quercetin was a strong antioxidant material in this study. Additional studies should examine the effects of various antioxidants on environmental stress factors and oxidative stress.

REFERENCES

- Ai, Q. H., K. S. Mai, C. X. Zhang, W. Xu, Q. Y. Duan, B. P. Tan and Z. G. Liufu. 2004. Effects of dietary vitamin C on growth and immune response of Japanese seabass, *Lateolabrax japonicus*. *Aquaculture* 242:489-500.
- Al-Jassabi, S. 2005. Biochemical studies on the role of lycopene in the protection of mice against microcystin toxicity. *J. Chem. Ecol.* 21:143-148.
- Atencio, L., I. Moreno, Jos Á, A. I. Prieto, R. Moyano, A. Blanco and A. M. Cameán. 2009. Effects of dietary selenium on the oxidative stress and pathological changes in tilapia

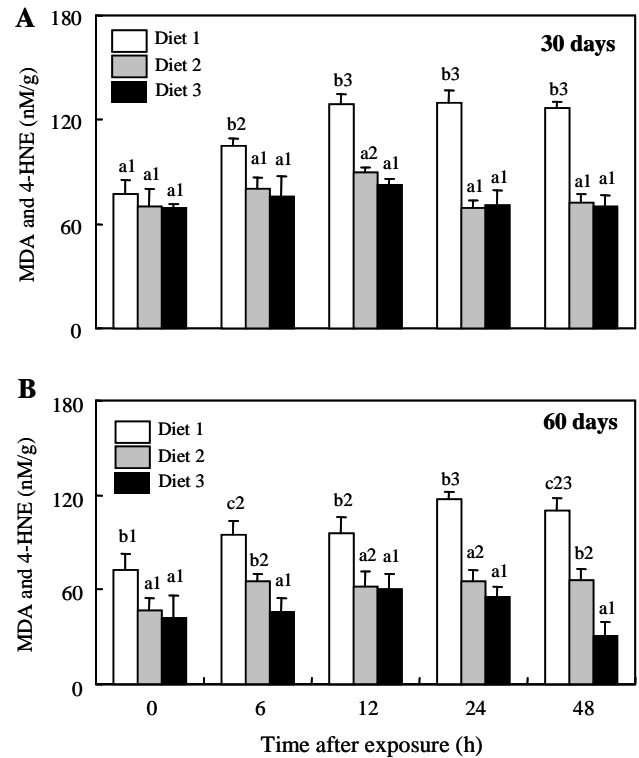


Figure 6. MDA and 4-HNE concentrations in the livers of olive flounder fed for 30 or 60 days diets containing quercetin 0.25 or 0.5%, after 6, 12, 24, and 48 h exposure to Cd. Values with alphabets indicate significant differences between Diet 1, 2 and 3 within the same time after exposure. The numbers indicate a significant difference from the control (within the same diet) ($p < 0.05$). All values are means \pm SD ($n = 5$).

(*Oreochromis niloticus*) exposed to a microcystin-producing cyanobacterial water bloom. *Toxicol* 53:269-282.

- Basha Siraj, P. and A. Rani Usha. 2003. Cadmium-induced antioxidant defense mechanism in freshwater teleost *Oreochromis mossambicus* (Tilapia). *Ecotoxicol. Environ. Saf.* 56:218-221.
- Bols, N. C., J. L. Brubacher, R. C. Ganassin and L. E. J. Lee. 2001. Ecotoxicology and innate immunity in fish. *Dev. Comp. Immunol.* 25:853-873.
- Bors, W. and M. Saran. 1987. Radical scavenging by flavonoid antioxidants. *Free Radic. Res. Commun.* 2:289-294.
- Chen, J., W. Q. Zhou, L. Feng, Y. Liu and J. Jiang. 2009. Effects of glutamine on hydrogen peroxide-induced oxidative damage in intestinal epithelial cells of Jian carp (*Cyprinus carpio* var. *Jian*). *Aquaculture* 288:285-289.
- da-Silva, W. S., J. W. Harney, B. W. Kim, J. Li, S. D. Bianco and A. Crescenzi. 2007. The small polyphenolic molecule kaempferol increases cellular energy expenditure and thyroid hormone activation. *Diabetes* 56:767-776.
- Dautremepuits, C., S. Betoulle, S. Paris-Palacios and G. Vernet. 2004. Humoral immune factors modulated by copper and chitosan in healthy or parasitized carp (*Cyprinus carpio* L.) by *tychobothrium* sp. (Cestoda). *Aquat. Toxicol.* 68:325-338.
- Eo, J. and K. J. Lee. 2008. Effect of dietary ascorbic acid on growth and non-specific immune responses of tiger puffer,

- Takifugu rubripes*. Fish Shellfish Immunol. 25:611-616.
- Esterbauer, H., R. J. Schaur and H. Zollner. 1991. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. Free Radic. Biol. Med. 11:81-128.
- Frankle, E. N., H. Kanner, J. B. German, E. Parks and J. E. Kinsella. 1993. Inhibition of oxidation of human low-density lipoprotein by phenolic substances in red wine. Lancet 341:454-457.
- Gouillou-Coustans, M. F., P. Bergot and S. J. Kaushik. 1998. Dietary ascorbic acid needs of common carp (*Cyprinus carpio*) larvae. Aquaculture 161:453-461.
- Hansen, B. H., S. Rømma, Ø. A. Garmo, P. A. Olsvik and R. A. Anderson. 2006. Antioxidative stress proteins and their gene expression in brown trout (*Salmo trutta*) from three rivers with different heavy metal levels. Comp. Biochem. Physiol. C 143:263-274.
- Hiratsuka, S., K. Ishihara, T. Kitagawa, S. Wada and H. Yokogoshi. 2008. Effect of dietary docosahexaenoic acid connecting phospholipids on the lipid peroxidation of the brain in mice. J. Nutr. Sci. Vitaminol. 54:501-506.
- Jayaraj, R., U. Deb, A. S. B. Bhaskar, G. B. K. S. Prasad and P. V. Lakshmana Rao. 2007. Hepatoprotective efficacy of certain flavonoids against microcystin induces toxicity in mice. Environ. Toxicol. 22:472-479.
- Kang, S-K., Y-D. Kim, K-H. Hyun, Y-W. Kim, J-S. Seo and Y-K. Park. 1998. Development of separating techniques on quercetin-related substances in onion (*Allium cepa* L.) 2. Optimal extracting condition of quercetin-related substances in onion. J. Korean Soc. Food Sci. Nutr. 27:687-692.
- Kinnula, V. L., J. D. Crapo and K. O. Raivio. 1995. Generation and disposal of reactive oxygen metabolites in the lung. Lab. Invest. 73:3-19.
- Merchie, G., P. Lavens, J. Verrth, F. Ollevier, H. Nelis, A. De Leenheer, V. Storch and P. Sorgeloos. 1997. The effect of supplemental ascorbic acid in enriched live food for *Clarias gariepinus* larvae at start feeding. Aquaculture 151:245-258.
- Monteiro, D. A., F. T. Rantin and A. L. Kalinin. 2009. The effects of selenium on oxidative stress biomarkers in the freshwater characid fish matrinxã (*Brycon cephalus*) (Günther, 1869) exposed to organophosphate insecticide Folisuper 600 BR (methyl parathion). Comp. Biochem. Physiol. C 149:40-49.
- Nouroozzadeh, J., J. Tajaddinisarmadi and S. P. Wolff. 1994. Measurement of plasma hydroperoxide concentrations by ferrous oxidation-xylenol orange assay in conjunction with triphenylphosphine. Anal. Biochem. 200:403-409.
- Ortuno, J., M. A. Esteban and J. Meseguer. 1999. Effect of high dietary intake of vitamin C on non-specific immune response of gilthead seabream (*Sparus aurata* L.). Fish Shellfish Immunol. 9:429-443.
- Pinho, G. L. L., C. Moura da Rosa, F. E. Maciel, A. Bianchini, J. S. Yunes, L. A. O. Proenca and M. J. Monserrat. 2005. Antioxidant responses after microcystin exposure in gills of an estuarine crab species pre-treated with vitamin E. Ecotoxicol. Environ. Saf. 61:361-365.
- Prieto, A. I., A. Jos, S. Pichardo, I. Moreno and A. M. Cameán. 2008. Protective role of vitamin E on the microcystin-induced oxidative stress in tilapia fish (*Oreochromis niloticus*). Environ. Toxicol. Chem. 27:1152-1159.
- Qinghui, A., K. Mai, C. Zhang, W. Xu, Q. Duan, B. Tan and Z. Liufu. 2004. Effects of dietary vitamin C on growth and immune response of Japanese seabass, *Lateolabrax japonicus*. Aquaculture 242:489-500.
- Scalbert, A. and G. Williamson. 2000. Dietary intake and bioavailability of polyphenols. J. Nutr. 130:2073S-2085S.
- Storey, K. B. 1996. Oxidative stress: animal adaptations in nature. Braz. J. Med. Biol. Res. 29:1715-1733.
- Trenzado, C. E., A. E. Morales, J. M. Palma and M. Higuera. 2009. Blood antioxidant defenses and hematological adjustments in crowded/uncrowded rainbow trout (*Oncorhynchus mykiss*) fed on diets with different levels of antioxidant vitamins and HUFA. Comp. Biochem. Physiol. C 149:440-447.
- Valavanidis, A., T. Vlahogianni, M. Dassenakis and M. Scoullas. 2006. Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants. Ecotoxicol. Environ. Saf. 64:178-189.
- Velioglu, Y. S. and G. Mazza. 1991. Characterization of flavonoids in peals of Rosa damascena by HPLC and spectral analysis. J. Agric. Food Chem. 39:463-467.
- Wang, W. and A. Li. 1996. Influences of LAPP on the growth, resistance to low oxygen content and immunoresistance of *Penaeus chinensis*. Trans. Oceanol. Limnol. 1:42-49.
- Weng, D., Y. Lu, Y. Wei, Y. Liu and P. Shen. 2007. The role of ROS in microcystin-LR-induced hepatocyte apoptosis and liver injury in mice. Toxicology 232:15-23.
- Wheeler, C. R., J. A. Salzman and N. M. Elsayed. 1990. Automated assays for superoxide dismutase, catalase, glutathione peroxidase, and glutathione reductase activity. Anal. Biochem. 184:193-199.
- Xu, C., W. Shu, Z. Qui, J. Chen, Q. Zhao and J. Cao. 2007. Protective effects of green tea polyphenols against subacute hepatotoxicity induced by microcystin-LR in mice. Environ. Toxicol. Pharmacol. 24:140-148.
- Zheng, Z. L., J. Y. W. Tan, H. Y. Liu, X. H. Zhou, X. Xiang and K. Y. Wang. 2009. Evaluation of oregano essential oil (*Origanum heracleoticum* L.) on growth, antioxidant effect and resistance against *Aeromonas hydrophila* in channel catfish (*Ictalurus punctatus*). Aquaculture 292:214-218.